

Supplementary Information

Derlin-1 is a rhomboid pseudoprotease required for the dislocation of mutant α -1-antitrypsin from the endoplasmic reticulum

Ethan J. Greenblatt^{1,2}, James A. Olzmann², and Ron R. Kopito^{1,2}

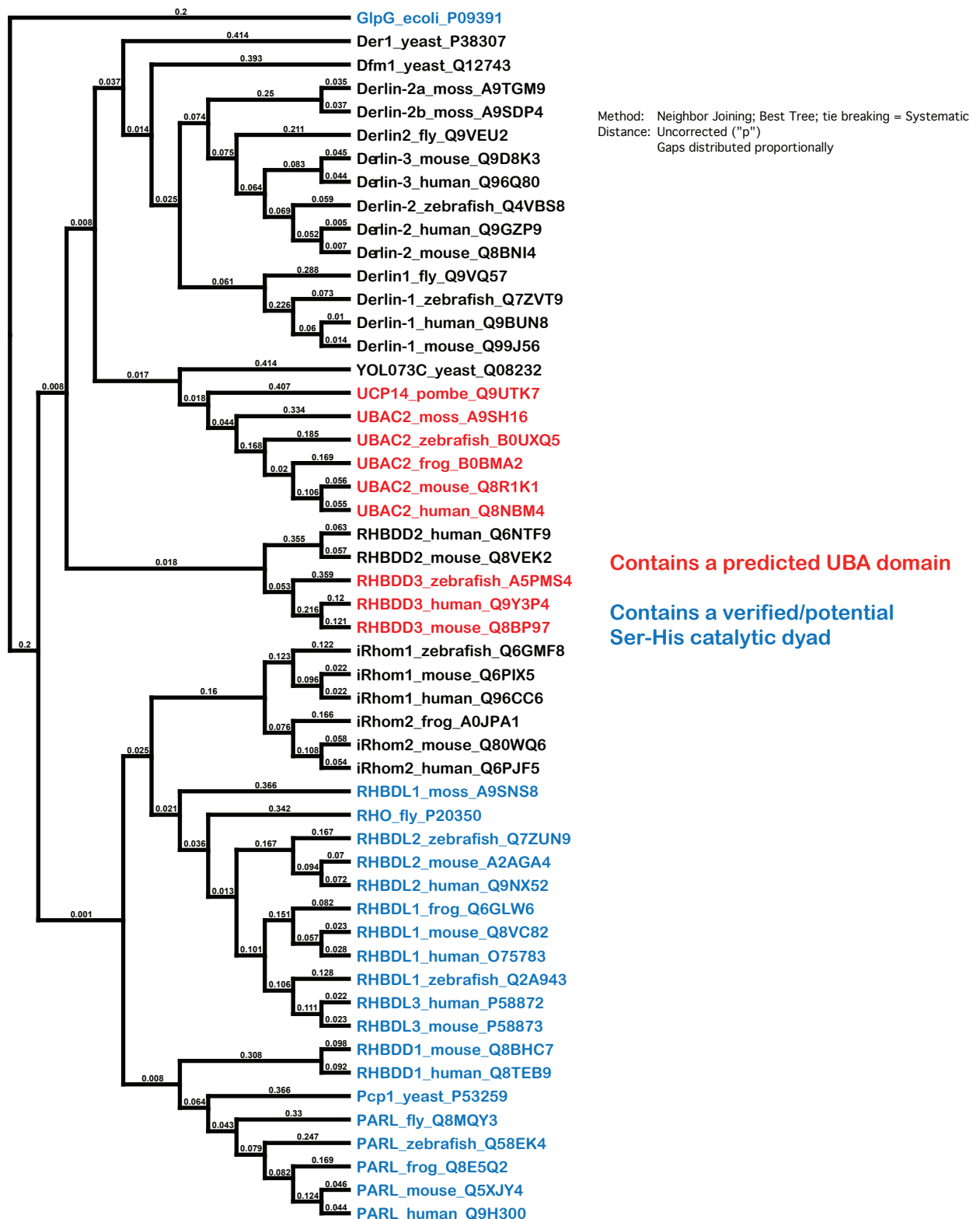
¹Biophysics Program, Stanford University, Stanford, California, USA

²Department of Biology, Stanford University, Stanford, California, USA

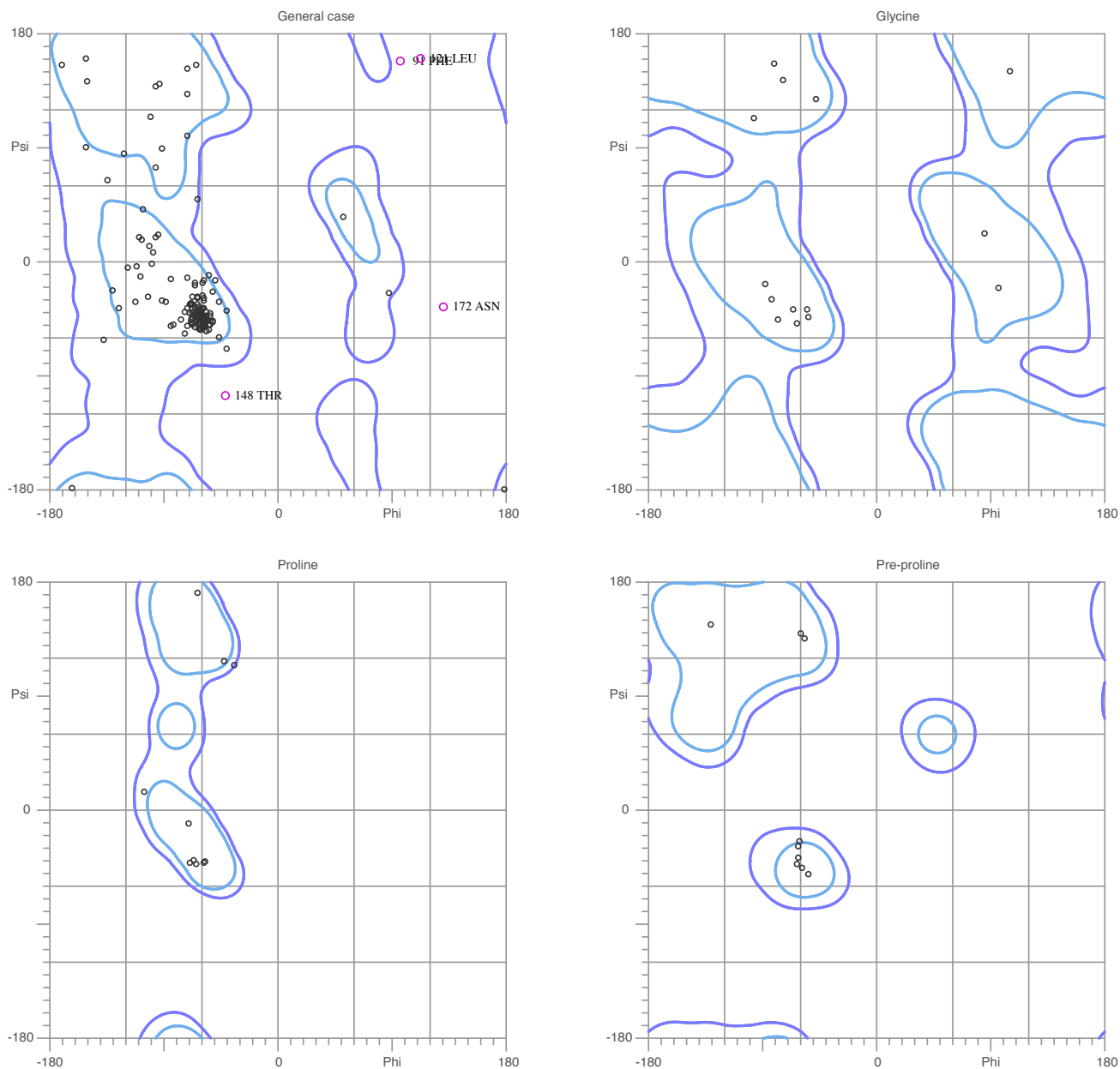
Supplementary information comprises:

Supplementary Figures 1–6

Supplementary References

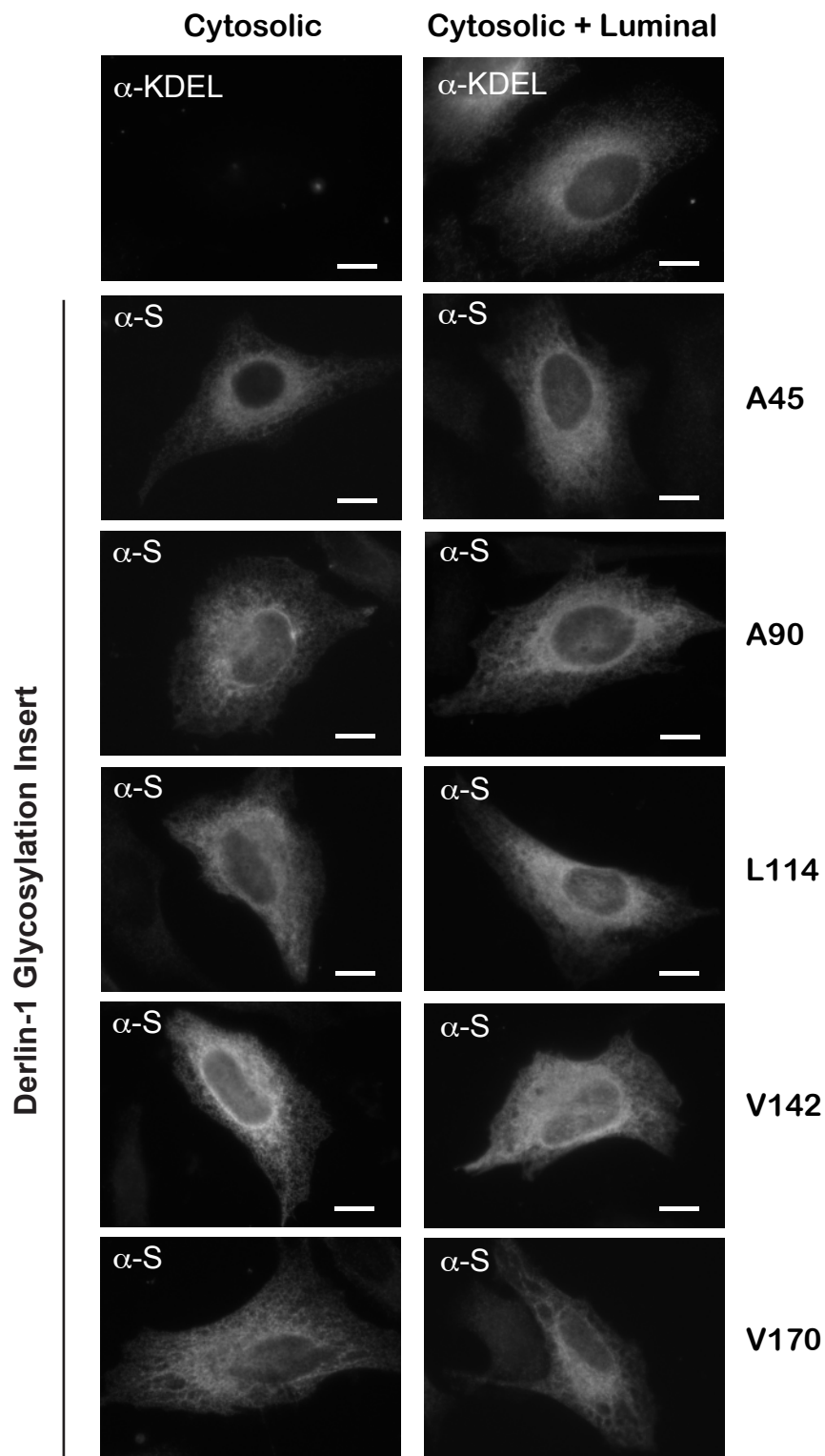


Supplementary Figure 1. Phylogenetic analysis of rhomboid family proteins. Sequences of eukaryotic rhomboid family proteins with the indicated UniProtKB accession numbers were obtained from the NCBI RefSeq database² and aligned using the multiple sequence and structure alignment server PROMALS3D. A phylogram was constructed by neighbor-joining using MacVector software with *E. coli* GlpG serving as an outgroup.

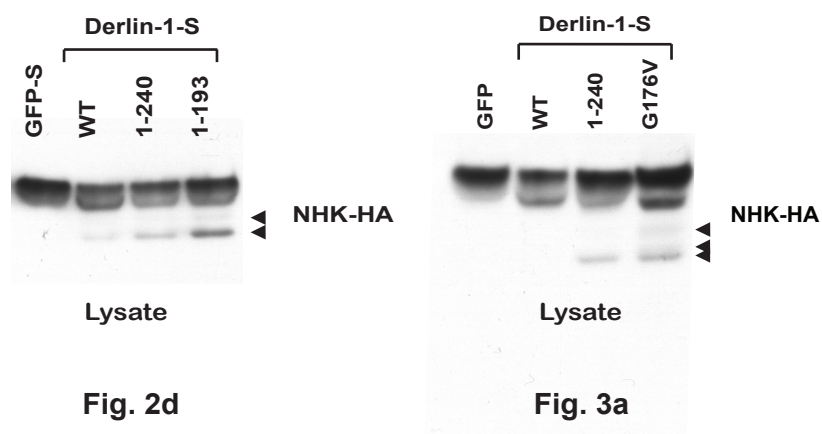


90.4% (161/178) of all residues were in favored (98%) regions.
 97.8% (174/178) of all residues were in allowed (>99.8%) regions.

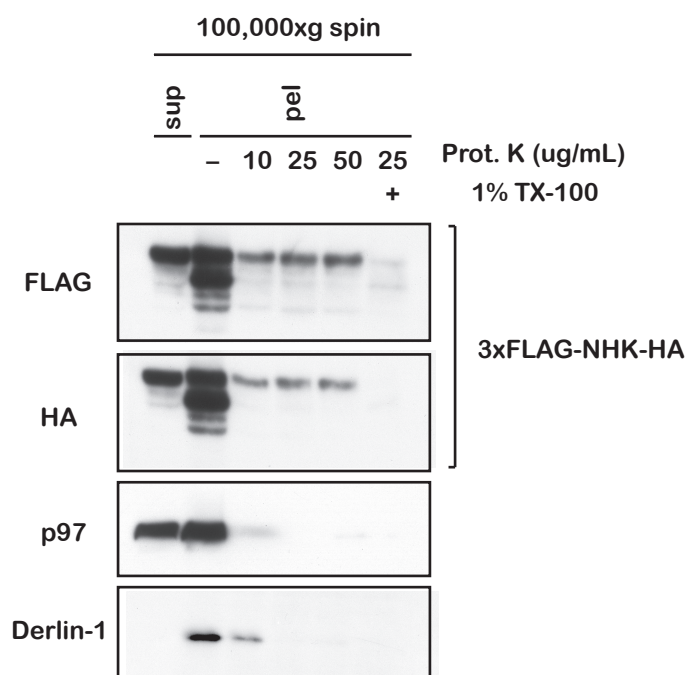
Supplementary Figure 2. Validation of the Derlin-1 homology model. Shown is the output of Ramachandran plot analysis by MolProbity of the Derlin-1 homology model depicted in Fig. 1b–d. The four outliers, F91, L121, T148, and N172 are each found in disordered loop regions connecting transmembrane spans.



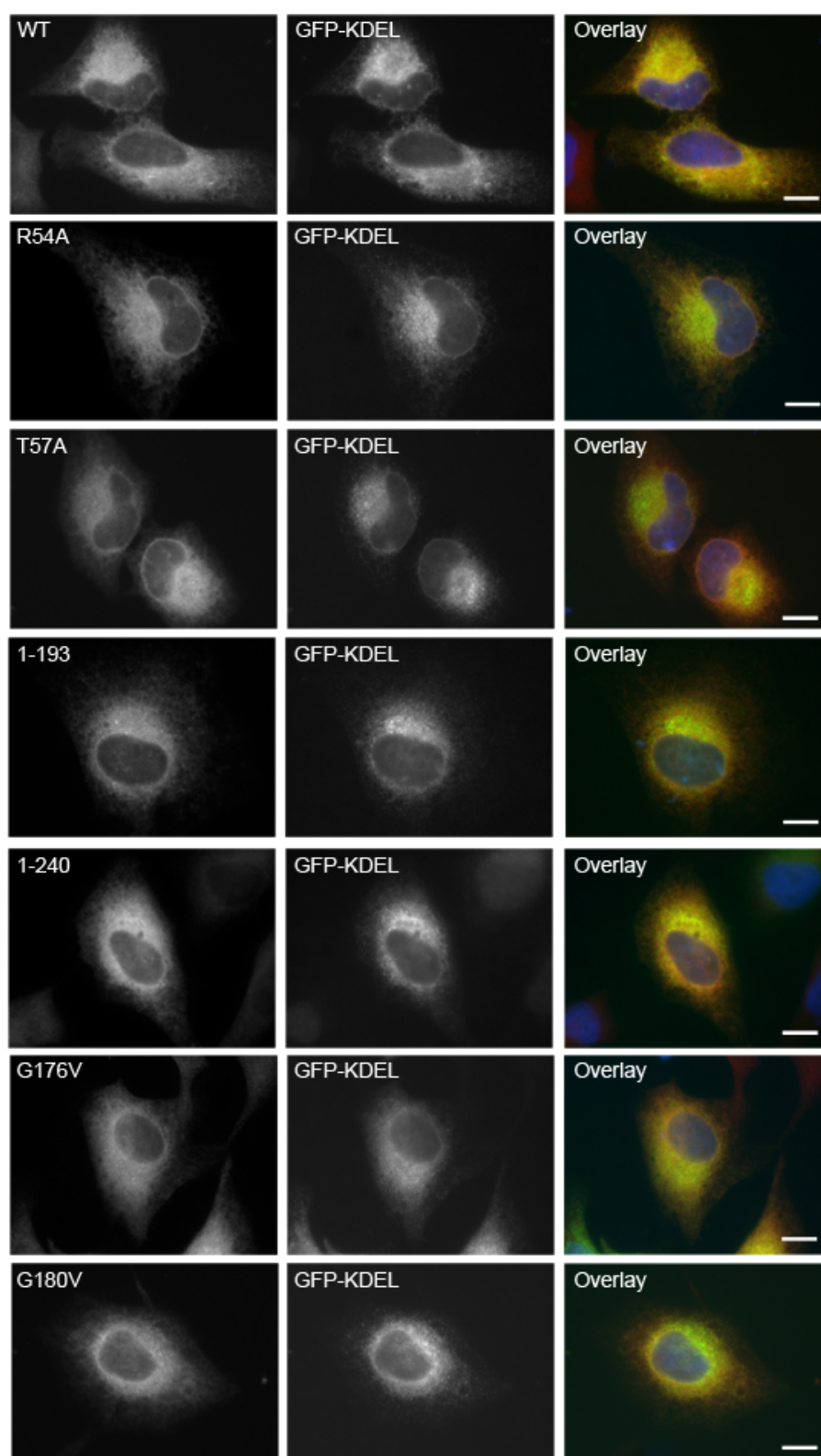
Supplementary Figure 3. Insertion of a 35 amino acid N-glycosylation acceptor sequence does not perturb the overall topology of Derlin-1. HeLa cells were transfected with the indicated chimeric Derlin-1 constructs used in Fig. 1g. Cells were treated with 20 μ M digitonin to permeabilize the plasma membrane only (Cytosolic) or with 20 μ M digitonin + 0.1% Triton X-100 to additionally permeabilize the ER (Cytosolic + Luminal), and stained with antibodies against KDEL or against S-tag, targeting C-terminally tagged Derlin-1-S.



Supplementary Figure 4. Light exposures of immunoblots from Fig. 2d and Fig. 3a. The shorter-length exposures of immunoblots from the indicated figures are shown above. Arrowheads indicate deglycosylated NHK-HA.



Supplementary Figure 5. Expression of Derlin-1_{G176V} stabilizes NHK on the cytoplasmic face of the ER. Membranes from cells co-expressing S-tagged Derlin-1_{G176V} and N- and C- terminally tagged 3xFLAG-NHK-HA were collected and proteolyzed as in Fig. 3c. In addition, the supernatant was collected following homogenization of cells and 100,000g centrifugation. Proteins in this fraction, which include cytosolic as well as non-membrane-associated luminal ER proteins released during homogenization, were precipitated with 15% TCA. Equal fractions of supernatant and membrane samples were analyzed by immunoblotting.



Supplementary Figure 6. S-tagged wild-type and mutant Derlin-1 localize properly to the endoplasmic reticulum. The indicated S-tagged Derlin-1 variants were co-expressed with the ER marker GFP-KDEL¹ in HeLa cells. Cells were analyzed by immunofluorescence with antibodies against S-tag.

Supplementary References

- 1 Terasaki, M., Jaffe, L. A., Hunnicutt, G. R. & Hammer, J. A. Structural change of the endoplasmic reticulum during fertilization: evidence for loss of membrane continuity using the green fluorescent protein. *Dev Biol* **179**, 320-328 (1996).
- 2 Pruitt, K. D., Tatusova, T., Klimke, W. & Maglott, D. R. NCBI Reference Sequences: current status, policy and new initiatives. *Nucleic Acids Res* **37**, D32-36, doi:10.1093/nar/gkn721 (2009).
- 3 Pei, J., Tang, M. & Grishin, N. V. PROMALS3D web server for accurate multiple protein sequence and structure alignments. *Nucleic Acids Res* **36**, W30-34, doi:10.1093/nar/gkn322 (2008).